

Galaxy Publication

Influence of Alpha Thalassemia and Hemoglobin F Co-inheritance on Sickle Cell Disease Outcomes

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ABSTRACT

The presence of alpha thalassemia and levels of hemoglobin F (HbF) significantly influence the clinical presentation of sickle cell disease (SCD) across different populations. This study was conducted to investigate the effects of these two factors on SCD patients in northern Iraq. A total of 74 patients with sickle/ β 0 thalassemia or sickle cell anemia, with a mean age of 16 years, participated in the study, of which 56.8% were male. Comprehensive clinical evaluations and lab tests were performed, including blood and reticulocyte counts, HbF levels, and analysis of serum lactic dehydrogenase and bilirubin. Screening for alpha-thalassemia mutations was performed using multiplex PCR and reverse hybridization. The results showed a positive correlation between HbF levels and hemoglobin, as well as negative correlations with reticulocyte count, HbA2, and the frequency of blood transfusions (P = 0.033, 0.041, 0.037, and 0.02, respectively). However, HbF was not correlated with other clinical symptoms. Nine patients had alpha-thalassemia (eight with $-\alpha 3.7/\alpha \alpha$ and one with $-\alpha 4.2/\alpha \alpha$), but no significant hematological or clinical effects were observed in these patients. This study showed that HbF, rather than alpha-thalassemia, primarily modulates the disease phenotype in SCD patients from northern Iraq, which is different from findings in populations from Africa, the Arabian Peninsula, and Iraq, where alpha-thalassemia often plays a more influential role, sometimes more than HbF.

Keywords: HbF, Sickle cell disease, Alpha thalassemia, Phenotype, Iraq

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Introduction

Sickle cell disease (SCD) is an inherited blood disorder caused by a mutation in the β -globin gene at codon 6, resulting in the production of hemoglobin S (HbS, $\alpha 2\beta S2$). This variant of hemoglobin tends to polymerize when deoxygenated, leading to red blood cells adopting a sickle shape [1]. This sickling reduces cell lifespan (hemolysis), impairs deformability, and causes difficulty navigating narrow blood vessels, which results in blockages and multi-organ damage [2]. SCD is most common among individuals of African descent, but it also affects populations in India, the Arabian Peninsula, and the Mediterranean [1]. In Iraq, SCD shows a distinct geographic distribution, being more frequent in the southern Basrah and northern Duhok provinces, while it is rare in other regions of the country [3]. The disease presents in various genetic forms, with the most common being sickle cell anemia (homozygosity for the sickle cell gene), followed by compound heterozygosity with β -thalassemia or other structural hemoglobinopathies, such as Hb SC or Hb SD [4]. The clinical progression of SCD

varies widely, making its course difficult to predict. Extensive research has been conducted to identify genetic modifiers that might help predict disease severity and offer potential therapeutic targets. Key factors influencing SCD include increased fetal hemoglobin (Hb F) production and the co-inheritance of α -thalassemia [5]. These factors' effects differ across populations, and this study aims to evaluate how they impact hematological and clinical outcomes in SCD patients from Northern Iraq.

Materials and Methods

Patients

Seventy-four patients diagnosed with sickle cell disease were enrolled in the study from the Inherited Blood Disorders Center in Duhok, Iraq. The study focused on confirmed cases of sickle cell anemia (SCA) and sickle/ β 0 thalassemia, excluding individuals with sickle/ β + thalassemia. Inclusion criteria required patients to be aged 2 years or older, have a confirmed SCD diagnosis, and be in a stable condition, defined as no sickle cell crises for at least four weeks. Patients who were on hydroxyurea treatment were not included. The study received approval from the ethics committee of the Health Directorate in Duhok, Iraq, and informed consent was obtained from all participants or their guardians.

Each participant underwent a comprehensive medical history review and physical examination, with a thorough assessment of their medical records. This included documenting demographic information and evaluating clinical events such as the occurrence of sickle cell crises, the number of blood transfusions, pain episodes, hospitalizations due to vaso-occlusive crises, and any other sickle cell-related admissions over the past year.

Upon enrollment, various tests were performed, including complete blood counts (Swelab, BouleMedical AB, Spånga, Sweden), reticulocyte counts, and high-performance liquid chromatography (HPLC) to measure HbF and HbA2 (D10, BioRad Laboratories, Hercules, CA, USA). Biochemical testing involved measuring serum bilirubin, ferritin, and lactate dehydrogenase (LDH) levels using an automated biochemistry analyzer (Cobas c501, Roche Diagnostics, HITACHI, Tokyo, Japan).

DNA was extracted using a modified salted-out technique that ensures high-quality and pure DNA yield [6]. Screening for alpha-thalassemia was carried out using multiplex polymerase chain reaction (PCR) and reverse hybridization with allele-specific oligonucleotide probes (ViennaLab Diagnostics GmbH, Vienna, Austria). This method detects 21 different α -globin mutations, including deletions such as $-\alpha 3.7$, $-\alpha 4.2$, --MED, --SEA, --THAI, --FIL, and $-(\alpha)20.5$, along with the $\alpha\alpha\alpha$ anti-3.7 gene triplication. It also identifies non-deletional α -thalassemias, such as mutations in the $\alpha 1$ gene (codon 14 (G > A) and Hb Adana (codon 59 (G > A)), and 11 point mutations on the $\alpha 2$ gene, including the initiation codon mutation ATG > ACG, codon 19 (-G), IVS-I, -5 nucleotides (-TGAGG), Hb Adana [codon 59 (G > A)], Hb Quong Sze [codon 125 (T > C)], Hb Constant Spring [codon 142 (T > C)], Hb Icaria [codon 142 (T > A)], Hb Paksé [codon 142 (A > T)], Hb Koya Dora [codon 142 (A > C)], and variations in polyadenylation signal sites (poly A1 [Saudi type] and poly A2 [Turkish type]).

Statistical Methods

Data analysis was conducted using SPSS software (Version 22, SPSS Corporation, Chicago, IL, USA). Descriptive statistics for continuous variables were expressed as median values with interquartile ranges (IQR). To compare groups, the Mann-Whitney test, the Kruskal-Wallis test, and Spearman correlation were applied as appropriate. A P-value < 0.05 was considered statistically significant.

Results and Discussion

The median age of the patients included in the study was 16.0 years (with a range of 2 to 47 years), comprising 42 males and 32 females. Among them, 59 had sickle cell anemia (SCA), and 15 had sickle/ β 0 thalassemia. **Table 1** presents the key clinical and laboratory characteristics of the enrolled patients.

Parameter	Median IQR		
Age (years)	16.0	10-23	
Sex (Male: Female)	42: 32		
Hb (g/L)	88	79–98	
Reticulocyte (%)	12.5	8-15	

 Table 1. Key clinical and laboratory characteristics of 74 patients with sickle cell disease

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MCV (fL)	84.2	75.8–91.7	
MCH (pg)	28.4	25.1-32.0	
WBC (x10^9/L)	12.9	9.6-16.9	
Platelets $(x10^{9}/L)$	440.0	251-616	
Bilirubin (µmol/L)	58.1	41.9-94.6	
HbF (%)	11.4	6.7-20.1	
HbA2 (%)	3.15	2.78-3.85	
LDH (IU/L)	555.0	304–934	
Ferritin (ng/mL)	155.1	92-382	
Overall hospitalization per year	1.0	0–3	
Hospitalization for VOC per year	1.0	0–2	
Pain episodes per year	2.0	0–6	
Blood transfusions per year	0	0–1.25	

History of Conditions

- Acute Chest Syndrome: 13 (17.6%)
- Splenectomy: 13 (17.6%)
- Avascular necrosis of femoral head: 7 (9.5%)
- Aplastic Crisis: 4 (5.4%)
- Splenic Sequestration: 4 (5.4%)
- Leg ulcers: 1 (1.4%)

IQR: Interquartile range

Correlation of Hemoglobin F (%) with Clinical and Laboratory Variables

The relationship between hemoglobin F (%) and various clinical and laboratory parameters was examined, with the results outlined in **Table 2**. The main findings revealed a positive correlation between Hemoglobin F and hemoglobin levels (**Figure 1**) as well as RBC counts (P = 0.033 and 0.009, respectively). On the other hand, negative correlations were observed with reticulocyte count (**Figure 2**), Hb A2 (%), frequency of blood transfusions, and platelet count (P = 0.041, 0.037, 0.020, and 0.030, respectively). Additionally, no significant association was found between Hemoglobin F and the yearly incidence of pain episodes or hospitalizations. It also showed no significant differences between patients with or without a history of sickle cell crises.

Parameter	HbF	Spearman coefficient	P-value	Mann-Whitney test (P Value)
Age	-0.131	0.265		
Hb	0.248	0.033*		
RBC count	0.303	0.009*		
MCV	-0.108	0.360		
MCH	-0.177	0.131		
Reticulocyte count	-0.238	0.041*		
WBC count	-0.109	0.354		
Platelet count	-0.253	0.030*		
Hb A2	-0.242	0.037*		
LDH	0.016	0.894		
S. Ferritin	-0.003	0.980		
S. Bilirubin	-0.142	0.227		
Frequency of blood transfusions per year	-0.270	0.020*		
Overall hospitalization per year	0.051	0.615		
Hospitalization for VOC per year	0.124	0.294		
Pain episodes per year	0.125	0.289		
Sex				0.639
Acute chest syndrome				0.972
Splenectomy				0.842
Avascular necrosis of the femoral head				0.305
Splenic sequestration				0.311
Aplastic crisis				0.990

 Table 2. Non-parametric correlations and associations between hb f (%) and various hematological and clinical variables in 74 patients with sickle cell disease

Note: Asterisks indicate statistically significant correlations.

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Figure 1. A scatterplot showing the positive correlation between HbF and hemoglobin concentration (Spearman coefficient = 0.248; P = 0.033)



Figure 2. A scatterplot showing the negative correlation between HbF and the reticulocyte count (Spearman coefficient -0.238, P = 0.041)

Alpha Thalassemia and Clinical Characteristics

Out of the patients studied, nine were diagnosed with alpha thalassemia, including eight with the $(-\alpha 3.7/\alpha \alpha)$ mutation and one with $(-\alpha 4.2/\alpha \alpha)$. No cases of double α -gene deletions or non-deletional α -thalassemia were detected. When comparing those with and without α -thalassemia, no significant differences in clinical or laboratory parameters were found (**Table 3**). Interestingly, none of the nine α -thalassemia carriers had a history of avascular necrosis of the femoral head, splenic sequestration, aplastic crisis, or leg ulcers, and only one individual had a history of acute chest syndrome (all P > 0.05).

Further analysis categorized patients into SCA, sickle/ β 0-thalassemia, and HbSS/ α -thalassemia groups (**Table 4**). The results showed that Hemoglobin F levels were significantly higher in sickle/ β 0-thalassemia, followed by SCA, and then HbSS/ α -thalassemia (P = 0.032). Hb A2 levels were highest in sickle/ β 0-thalassemia, then HbSS/ α -thalassemia, and lowest in SCA (P = 0.001). In terms of MCV and MCH, sickle/ β 0-thalassemia exhibited the lowest values, followed by HbSS/ α -thalassemia, with SCA showing the highest (P < 0.001 for both). Additionally, the frequency of blood transfusions was most frequent in SCA, followed by HbSS/ α -thalassemia, and least in sickle/ β 0-thalassemia (P = 0.039).

 Table 3. Comparison between various continuous variables in sickle cell disease with or without α-thalassemia using Mann-Whitney U test

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Parameter	Media	Davalaa		
rarameter	SCD without α-thal	SCD with a-thal	– P-value	
Age (years)	16 (10-23)	15 (10.5-25.5)	0.993	
Hb (g/L)	88 (79.5-98)	86 (76-100)	0.766	
RBC (x10 ¹² /L)	3.04 (2.5-3.7)	3.05 (2.9-3.3)	0.960	
Reticulocyte count (%)	12 (8.0-15.0)	13.0 (10.1-15.0)	0.476	
MCV (fL)	84.3 (75.6-94)	79.2 (74.9-88.2)	0.350	
MCH (pg)	28.4 (24.9-33.1)	26.4 (25.5-29.9)	0.376	
HbF (%)	13.6 (7.3-20.95)	7.4 (5.213.1)	0.077	
HbA2 (%)	3.1 (2.65-4.0)	3.5 (3-3.75)	0.466	
LDH (IU/L)	540 (297-917)	570 (422-1204)	0.602	
Ferritin (ng/ml)	152 (83-363)	162 (103-538)	0.710	
Overall hospitalization annual	1 (0-3)	0 (0-2)	0.311	
Hospitalization for VOC (Annual)	1 (0-2)	0 (0-1)	0.182	
Pain episodes annual	2 (0-6)	0 (0-2)	0.092	
Blood transfusion annual	0 (0-1.5) 0 (0-1.5)		0.837	
Bilirubin (umol/L)	56.4 (39.3-95.1) 59.9 (47-93.2)		0.882	
WBC (x10 ⁹ /L)	12.6 (9.1-17.0)	14.1 (11.8-17.2)	0.350	
Platelets (x10 ⁹ /L)	438 (248-618)	489 (269-581)	0.862	

*VOC: Vaso-occlusive crisis.

Table 4. Comparison between various continuous variables in sickle cell anemia (SCA), sickle/ β^0 -thalassemia, and Hb SS/ α -thalassemia using the Kruskal-Wallis test

Parameter	Median (IQR)				
	SS/α-thal	Sickle/β ⁰ thal	SCA	- P-value	
Age (years)	15 (10.5-25.5)	16 (10-25)	16.5 (9.8-23)	0.896	
Hb (g/L)	86 (76-100)	95 (80-99)	86 (77-97)	0.445	
RBC (x10 ¹² /L)	3.05 (2.9-3.3)	4.0 (3.7-4.3)	2.86(2.40-3.45)	< 0.001	
Reticulocyte count (%)	13.0 (10.1-15.0)	14.0 (9-15)	11.7 (8-15)	0.587	
MCV (fL)	79.2 (74.9-88.2)	71.8 (68.7-73.4)	86.4 (83-95.9)	< 0.001	
MCH (pg)	26.4 (25.5-29.9)	23.5 (22.7-24.2)	30.0 (27.4-33.6)	< 0.001	
HbF (%)	7.4 (5.213.1)	21.2 (8.2-28.1)	12.9 (6.7-18.8)	0.032	
HbA2 (%)	3.5 (3-3.75)	4.8 (3.1-5.4)	3.0 (2.5-3.4)	0.001	
LDH (IU/L)	570 (422-1204)	700 (446-1107)	521 (278-880)	0.140	
Ferritin (ng/ml)	162 (103-538)	181 (102-350)	146 (71-382)	0.661	
Overall hospitalization annual	0 (0-2)	1 (0-2)	1 (0-4.25)	0.369	
Hospitalization for VOC (Annual)	0 (0-1)	0 (0-2)	1 (0-2.25)	0.283	
Pain episodes annual	0 (0-2)	4 (2-7)	2 (0-6)	0.115	
Blood transfusion annual	0 (0-1.5)	0 (0-0)	0 (0-2.25)	0.039	
Bilirubin (umol/L)	59.9 (47-93.2)	44.5 (34.2-56.4)	63.3 (46.2-99.2)	0.043	
WBC (x10 ⁹ /L)	14.1 (11.8-17.2)	12.6 (9.1-17.0)	12.7 (9.1-16.9)	0.645	
Platelets (x10 ⁹ /L)	489 (269-581)	438 (181-580)	438 (251-638)	0.895	

*VOC: Vaso-occlusive crisis

Red Cell Indices and Their Correlations with Laboratory and Clinical Parameters

The relationships between various red cell indices, laboratory measurements, and clinical variables were explored, with some significant correlations summarized in Table 5. Notable findings include: a negative correlation between hemoglobin levels and both reticulocyte count and LDH (P = 0.001 and 0.022, respectively); LDH showing a positive correlation with reticulocyte count, pain episode frequency, and transfusion frequency (P = 0.007, 0.030, and 0.006, respectively); and both MCV and MCH being positively correlated with transfusion frequency (P = 0.034 and 0.020, respectively). Further correlations are provided in **Table 5**.

Table 5. Some significant bivariate correlations between various hematological and clinical parameters in 74

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Parameters	Spearman coefficient	P-value	
Hemoglobin and age	+ 0.431	< 0.001	
Hemoglobin and RBC count	+0.666	< 0.001	
Hemoglobin and reticulocyte count	- 0.389	0.001	
Hemoglobin and Hb F (%)	+0.284	0.033	
Hemoglobin and LDH	-0.266	0.022	
Hemoglobin and platelets	+0.318	0.006	
Reticulocyte count and RBC count	-0.318	0.006	
Reticulocyte count and Hb F(%)	-0.238	0.041	
Reticulocyte count and LDH	0.309	0.007	
Hb F and RBC count	0.303	0.009	
Hb F and transfusion frequency/year	-0.270	0.020	
Hb F (%) and Hb A2 (%)	-0.242	0.037	
Hb F(%) and platelets	-0.253	0.030	
LDH and frequency of pain episodes/year	+0.253	0.030	
LDH and frequency of transfusions/year	+0.318	0.006	
Frequency of hospitalization and pain episodes /year	+0.688	< 0.001	
Frequency of hospitalization and transfusions/year	+0.272	0.019	
Leucocytes and frequency of hospitalization/year	+0.219	0.061	
Leucocytes and platelet counts	+0.374	0.001	

SCD disease patients were examined

Assessment of Clinical and Laboratory Parameters in Sickle Cell Disease

When evaluating the history of sickle cell crises about other laboratory and clinical parameters, several important associations were observed. Acute chest syndrome was linked with more frequent pain episodes and hospital admissions, though only the association with pain episodes was statistically significant (P = 0.037). Avascular necrosis of the femoral head showed no significant correlations with any tested parameters, although these patients tended to be slightly older (P = 0.102). In contrast, a history of splenectomy was significantly associated with older age, higher hemoglobin, and platelet counts (P = 0.020, 0.048, and 0.043, respectively).

In Iraq, sickle cell disease (SCD) presents with a distinct geographical distribution, with two primary epicenters: one in the north, associated with the Benin haplotype, and another in the south, linked to the Arab Indian haplotype. The Benin haplotype is associated with moderate disease severity, while the Arab Indian haplotype often corresponds with higher levels of HbF and a milder disease course. The current study's findings, including a median HbF of 11.4% and hemoglobin levels of 88 g/L, align with what has been observed in areas with the Benin haplotype, such as Southwest Saudi Arabia, but are lower than those in regions with the Arab Indian haplotype, such as Southern Iraq and the Arabian Peninsula [7-11].

Hemoglobin F (HbF) has been identified as a potent modulator of sickle cell disease phenotypes, as it inhibits the polymerization of deoxygenated HbS and reduces the mean concentration of HbS in red cells. In this study, HbF was positively correlated with hemoglobin concentration and negatively with reticulocyte count and transfusion frequency, which is consistent with its protective role. Similar findings have been reported in studies of Jamaican and American SCD patients [12-14]. Additionally, a negative correlation between HbF and HbA2 was observed, which may reflect a preferential survival of cells with higher HbF levels. Despite these correlations, no significant association between HbF and clinical manifestations like pain episodes or other sickle cell crises was found, aligning with previous research in Jamaican and Saudi Arabian populations. However, other studies have shown that higher HbF levels are linked to a reduced frequency of vaso-occlusive episodes [12, 15, 16].

Alpha thalassemia, which reduces α -chain production and subsequently lowers the intracellular HbS concentration, was found in 12.2% of the studied sample. This is similar to the rate observed in Northern Iraq but lower than in Southern Iraq or the Arabian Peninsula countries. The co-inheritance of α -thalassemia with sickle cell disease has been associated with various clinical outcomes, including higher hemoglobin levels, lower reticulocyte counts, and protection against certain complications like stroke and leg ulcers. However, the current study found no significant differences in hemoglobin or reticulocyte counts between those with or without α -thalassemia. Although fewer pain episodes were noted in patients with HbSS/ α -thalassemia, no significant clinical associations were found. This could be due to the predominance of single α -gene deletions in this study, while

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previous studies often included cases with double gene deletions, which are believed to have a more substantial ameliorating effect on SCD symptoms [17-33].

The study's limitations include its cross-sectional design, its focus on patients in a steady state, and the exclusion of individuals on hydroxyurea therapy, which may have reduced the pool of eligible participants.

Conclusion

In conclusion, the findings of this study indicate that in SCD patients from Northern Iraq, hemoglobin F (HbF) plays a more significant role in modulating the disease phenotype than α -thalassemia. This contrasts with the findings in many studies from Africa, the Arabian Peninsula, and Southern Iraq, where α -thalassemia is a major modulator of the disease and, in some cases, even has a greater effect than HbF.

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References

- 1. Piel FB, Steinberg MH, Rees DC. Sickle cell Anemia. N Engl J Med. 2017;376:1561-73.
- 2. Kavanagh PL, Fasipe TA, Wun T. Sickle cell disease. JAMA. 2022;328(1):57-68.
- 3. Al-Allawi N, Al Allawi S, Jalal S. Genetic epidemiology of hemoglobinopathies in Iraqi Kurds. J Community Genet. 2021;12(1):5-14.
- 4. Ballas SK, Lieff S, Benjamin LJ, Dampier CD, Heeney MM, Hoppe C, et al. Definitions of the phenotypic manifestations of sickle cell disease. Am J Hematol. 2010;85(1):6-13.
- 5. Thein SL. Genetic association studies in β-hemoglobinopathies. Hematology Am Soc Hematol Educ Program. 2013;2013(1):354-61.
- 6. Kashmoola MA, Eissa AA, Al-Takay DT, Al-Allawi NA. Molecular characterization of G6PD deficient variants in Nineveh Province, Northwestern Iraq. Indian J Hematol Blood Transfus. 2015;31(1):133-6.
- 7. Eissa AA, Markous RD, Yahya NB, Al-Allawi NA. Hemoglobin F modulation in sickle cell disease: Experience in a single center in Iraqi Kurdistan. J Appl Hematol. 2016;7(3):85-9.
- 8. Al-Allawi NA, Jalal SD, Nerwey FF, Al-Sayan GO, Al-Zebari SS, Alshingaly AA, et al. Sickle cell disease in the Kurdish population of northern Iraq. Hemoglobin. 2012;36(4):333-42.
- 9. Yaseen NT, Al-Mamoori HS, Hassan MK. Sickle β-globin haplotypes among patients with sickle cell anemia in Basra, Iraq: A cross-sectional study. Iraqi J Hematol. 2020;9(1):23-9.
- 10. Loggetto SR. Sickle cell anemia: Clinical diversity and beta S-globin haplotypes. Rev Bras Hematol Hemoter. 2013;35(3):155-7.
- 11. Alsultan A, Aleem A, Ghabbour H, AlGahtani FH, Al-Shehri A, Osman ME, et al. Sickle cell disease subphenotypes in patients from Southwestern Province of Saudi Arabia. J Pediatr Hematol/Oncol. 2012;34(2):79-84.
- 12. Steinberg MH, Sebastiani P. Genetic modifiers of sickle cell disease. Am J Hematol. 2012;87(8):795-803.
- 13. Maude GH, Hayes RJ, Serjeant GR. The haematology of steady state homozygous sickle cell disease: Interrelationships between haematological indices. Br J Haematol. 1987;66(4):549-58.
- 14. Odenheimer DJ, Whitten CF, Rucknagel DL, Sarnaik SA, Sing CF. Heterogeneity of sickle-cell anemia based on a profile of hematological variables. Am J Hum Genet. 1983;35(6):1224-40.
- 15. Donaldson A, Thomas P, Serjeant BE, Serjeant GR. Foetal haemoglobin in homozygous sickle cell disease: A study of patients with low HBF levels. Clin Lab Haematol. 2001;23(5):285-9.
- 16. Pembrey ME, Wood WG, Weatherall DJ, Perrine RP. Fetal haemoglobin production and the sickle gene in the oases of Eastern Saudi Arabia. Br J Haematol. 1978;40(3):415-29.

- Alsultan A, Alabdulaali MK, Griffin PJ, AlSuliman AM, Ghabbour HA, Sebastiani P, et al. Sickle cell disease in Saudi Arabia: The phenotype in adults with the Arab-Indian haplotype is not benign. Br J Haematol. 2014;164(4):597-604.
- Wonkam A, Mnika K, Ngo Bitoungui VJ, Chetcha Chemegni B, Chimusa ER, Dandara C, et al. Clinical and genetic factors are associated with pain and hospitalisation rates in sickle cell anaemia in Cameroon. Br J Haematol. 2018;180(1):134-46.
- 19. Jit BP, Mohanty PK, Purohit P, Das K, Patel S, Meher S, et al. Association of fetal hemoglobin level with frequency of acute pain episodes in sickle cell disease (HbS-only phenotype) patients. Blood Cells Mol Dis. 2019;75:30-4.
- 20. Tarer V, Etienne-Julan M, Diara JP, Belloy MS, Mukizi-Mukaza M, Elion J, et al. Sickle cell anemia in Guadeloupean children: pattern and prevalence of acute clinical events. Eur J Haematol. 2006;76(3):193-9.
- 21. Pandey S, Pandey S, Mishra RM, Sharma M, Saxena R. Genotypic influence of α-deletions on the phenotype of Indian sickle cell anemia patients. Korean J Hematol. 2011;46(3):192-5.
- 22. Platt OS, Thorington BD, Brambilla DJ, Milner PF, Rosse WF, Vichinsky E, et al. Pain in sickle cell disease: Rates and risk factors. N Engl J Med. 1991;325(1):11-6.
- Ballas SK. Effect of α-globin genotype on the pathophysiology of sickle cell disease. Pediatr Pathol Mol Med. 2001;20(2):107-21.
- Al-Barazanchi ZA, Abdulateef SS, Hassan MK. Co-Inheritance of α-thalassemia gene mutation in patients with sickle cell Disease: Impact on clinical and hematological variables. Niger J Clin Pract. 2021;24(6):874-82.
- Hassan SM, Al Muslahi M, Al Riyami M, Bakker E, Harteveld CL, Giordano PC. Sickle cell anemia and αthalassemia: A modulating factor in homozygous HbS/S patients in Oman. Eur J Med Genet. 2014;57(11-12):603-6.
- 26. Al-Ali AK, Alsulaiman A, Alfarhan M, Safaya S, Vatte CB, Albuali WM, et al. Sickle cell disease in the eastern province of Saudi Arabia: Clinical and laboratory features. Am J Hematol. 2021;96(4):E117-21.
- 27. Abuamer S, Shome DK, Jaradat A, Radhi A, Bapat JP, Sharif KA, et al. Frequencies and phenotypic consequences of association of α -and β -thalassemia alleles with sickle-cell disease in Bahrain. Int J Lab Hematol. 2017;39(1):76-83.
- 28. Steinberg MH, Emburyz SH. α-Thalassemia in blacks: Genetic and clinical aspects and interactions with the sickle hemoglobin gene. Blood. 1986;68(5):985-90.
- 29. Sheehan VA, Luo Z, Flanagan JM, Howard TA, Thompson BW, Wang WC, et al. Genetic modifiers of sickle cell anemia in the BABY HUG cohort: Influence on laboratory and clinical phenotypes. Am J Hematol. 2013;88(7):571-6.
- Saraf SL, Akingbola TS, Shah BN, Ezekekwu CA, Sonubi O, Zhang X, et al. Associations of α-thalassemia and BCL11A with stroke in Nigerian, United States, and United Kingdom sickle cell anemia cohorts. Blood Adv. 2017;1(11):693-8.
- Darbari DS, Onyekwere O, Nouraie M, Minniti CP, Luchtman-Jones L, Rana S, et al. Markers of severe vasoocclusive painful episode frequency in children and adolescents with sickle cell anemia. J Pediatr. 2012;160(2):286-90.
- 32. Mukherjee MB, Colah RB, Ghosh K, Mohanty D, Krishnamoorthy R. Milder clinical course of sickle cell disease in patients with α thalassemia in the Indian subcontinent. Blood J Am Soc Hematol. 1997;89(2):732.
- 33. Dover GJ, Chang VT, Boyer SH, Serjeant GR, Antonarakis SE, Higgs DR. The Cellular basis for different fetal hemoglobin levels among sickle cell individuals with two, three and four α -globin genes. Blood. 1987;69:341-4.